

## Human Babesiosis and Travelers' Health: Clinical and Public Health Issues

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### Abstract

Babesiosis is an infection caused by protozoa of the *Babesia* genus, whose vector are ticks of the *Ixodes scapularis* species. The disease is endemic in the United States, and, less frequently, in Europe; however, other regions occasionally may be affected. In non-endemic regions, like Brazil, the most of cases are associated with endemic areas travel. Thereby, the diagnosis should be proposed for patients with suggestive infection symptoms. Therefore, this review describes the illness caused by *Babesia* spp. and emphasizes the travelers' health aspects. The discussion was based on the Cochrane, Lilacs, Pubmed, Scielo, and Scielo Brasil databases, including the citations available up to May 30, 2017. Traveler awareness about the possible transmittable illnesses at the destination, specifically babesiosis in the United States and Europe is fundamental.

**Keywords:** Babesiosis; Protozoan infections; Traveler health

### Introduction

Babesiosis is an intraerythrocytic infection caused by protozoa of several species included in the genus *Babesia*. The pathogen is capable of infecting several animal groups naturally, including mammals, birds, reptiles, and amphibians [1,2] and humans are considered accidental hosts [3]. Transmission occurs by the bite of ticks belonging to the *Ixodidae* family, previously infected by the protozoa. In addition to this form of transmission, there are also cases caused by blood and congenital pathways [1,2].

In the United States and Europe, human babesiosis is generally associated with infection by *Babesia microti* and *Babesia divergens*, respectively [3]. However, *Babesia venatorum* emerged as a cause of the disease in China [4]. The first record of infection in the *Homo sapiens* species occurred in Yugoslavia in 1956 [5], and only three decades later Brazil reported the first case of the disease, more specifically in the state of Pernambuco [6], with further cases after this one uncommon in the country. However, the number of occurrences of human ixodidiosis has been increasing, accompanied by reports caused by agents transmitted by ticks [7,8].

The disease constitutes an occupational hazard for farmers, rangers, landscapers, hunters, and professionals who work in direct contact with the soil, road and railroad maintenance, and lumberjacking [9]. Travelers are of concern for health authorities in terms of babesiosis when considering the destination and duration of travel and the risk of exposure to infection [10]. It is important to point out that currently more than half a billion people are involved in international travel annually, which demonstrates the individual and collective risks arising from the movement of people

and their interaction with the various environments [10,11]. Noting the zoonotic and economic importance of babesiosis-with a special focus on travelers' health-this review presents the principal aspects related to etiology, life cycle, epidemiology, and clinical aspects of illness caused by *Babesia* spp.

### Etiology

Protozoa of the genus *Babesia* are included in the phylum *Apicomplexa*, of the suborder *Piroplasmida* and family *Babesiidae* [12]. They measure between 1 and 5 micrometers, and are oval, round or pyriform in shape. They present annular conformation and peripheral location, similar to the protozoa of the genus *Plasmodium falciparum*, which sometimes complicates diagnosis and may result in misidentification [13]. The absence of hemozoin deposits on the ring, absence of banana-shaped gametocytes, and presence of tetrads distinguish *Babesia* from *Plasmodium* [14].

More than 100 species infect several wild and domesticated animals and few have been confirmed as causative agents in humans, which vary depending on the region, with *Babesia divergens*, in Europe (especially in France and England), *B. microti* in the United States [15], more recently *B. venatorum* in China [4], also previously reported in Europe [16]. Cases of infection by *Babesia duncani* were registered in California and in the state of Washington [17]. Although, *B. microti* is present in Europe, there are few cases caused by this agent on the continent, most likely because the vector (*Ixodes trianguliceps*) is not a human parasite [18].

The *Babesia* species are classified into two forms, small and large. The former, represented by *Babesia microti*, *Babesia duncani*, and *Babesia conradae*, vary between 1 and 3 µm in diameter. Their trophozoites divide

by two successive binary fissions, resulting in four merozoites, arranged in a tetrad [13,19]. The large forms include *Babesia bovis*, *Babesia canis*, and *Babesia odocoilei*, with diameters varying between 3 and 5  $\mu\text{m}$  [13] and division only occurring in two pairs of merozoites (“paired form”) [13,19].

### Life Cycle

In vertebrates, including humans, the most common form of transmission of *Babesia* spp. is by the bite of ticks belonging to the *Ixodidae* family [2,13]. The endemism of the vector *Ixodes scapularis*, in Canada and most Eastern states of North America [20] coincide with that of babesiosis caused by *B. microti* [14].

The life cycle of *I. scapularis* is made up of three stages, larva, nymph, and adult [14]. The infection of newly hatched vector larva by *Babesia* spp. normally occurs at the end of summer. The parasite may cross the intestinal epithelium of the vector, with successive migration by the hemolymph and salivary glands [20]. However, the protozoa remain dormant during the transition from the larval to the nymph phase. At the start of the following summer, during the tick bite-together with the saliva of the nymph infected by *B. microti*-injection of sporozoites occurs, which are released on the host skin. All developmental forms – larva, nymph, and adult – feed on human blood; however the nymph is the principal vector due to its small size and greater activity in the summertime [14].

Transmission requires 36 to 72 hours, given that the sporozoites are only generated after activation, because of dormancy and exposure of the tick to the host’s warm blood, and are not immediately available in the salivary glands [14]. Afterwards, the sporozoite penetrates directly into the erythrocytes (*B. bovis* and *B. divergens*) or initially in the lymphocytes, where replication and release of “daughter cells” occurs, which in turn invade the red blood cells (*B. microti*). The host cell membrane undergoes intussusception, forming a vacuole that gradually disintegrates. Upon penetrating the erythrocyte, the organism transforms into the trophozoite form, able of moving freely in the cytoplasm [13].

The next step in the cycle is binary division, asexual reproduction known as merogony. Depending on the type of trophozoite, there will be division into two or four merozoites with in the red blood cells [13,21]. The red blood cells lose integrity after the exit of the merozoite and the departure of this developmental form allows for the cycle to re-begin. In the blood, some trophozoites differentiate into male and female gametocytes, within the red blood cells [13].

During the tick’s blood meal, trophozoites, merozoites, and gametocytes are ingested. However, only the gametocytes survive the intestinal lumen of the tick, where sexual reproduction occurs. This is followed by penetration of the zygote into the cells of the intestinal endothelium and asexual reproduction (sporogony), which results in sporokinets. These reach various organs within the tick’s body by means of the hemolymph, including the ovaries and salivary glands, locations where sporogony occurs again, producing infective sporozoites that are inoculated together with saliva during the vector’s feeding [22]. Some species of *Babesia* spp. show transovarian transmission to vector for all phases of the life cycle [12].

Transmission of the protozoan to humans can also occur by congenital route or by blood transfusion [2,13]. The parasite can survive for up to 35 days at 4°C [23].

Transmission of *Babesia* sp. to humans occurs most frequently by the bite of the infected vector, but also occurs by blood transfusion. In this model (Figure 1), ticks infected by *Babesia* sp. introduce the sporozoites into the host during the bite, together with saliva (1). The sporozoites entre the red blood cells and reproduce asexually (2). In the blood, some parasites differentiate into male and female gametes (3). Upon ingestion by the tick, the gametes (4) unite forming new sporozoites (5). Humans enter the cycle accidentally, upon being bitten by ticks infected with *Babesia* sp., and consequent sporozoite injection (6), which penetrate the erythrocytes (B) and reproduce asexually (7). Blood transfusion is another form of transmission (8). Transovarian transmission occurs in the “large” forms of *Babesia* sp (A) (Source: Centers for Disease Control and Prevention (CDC)).

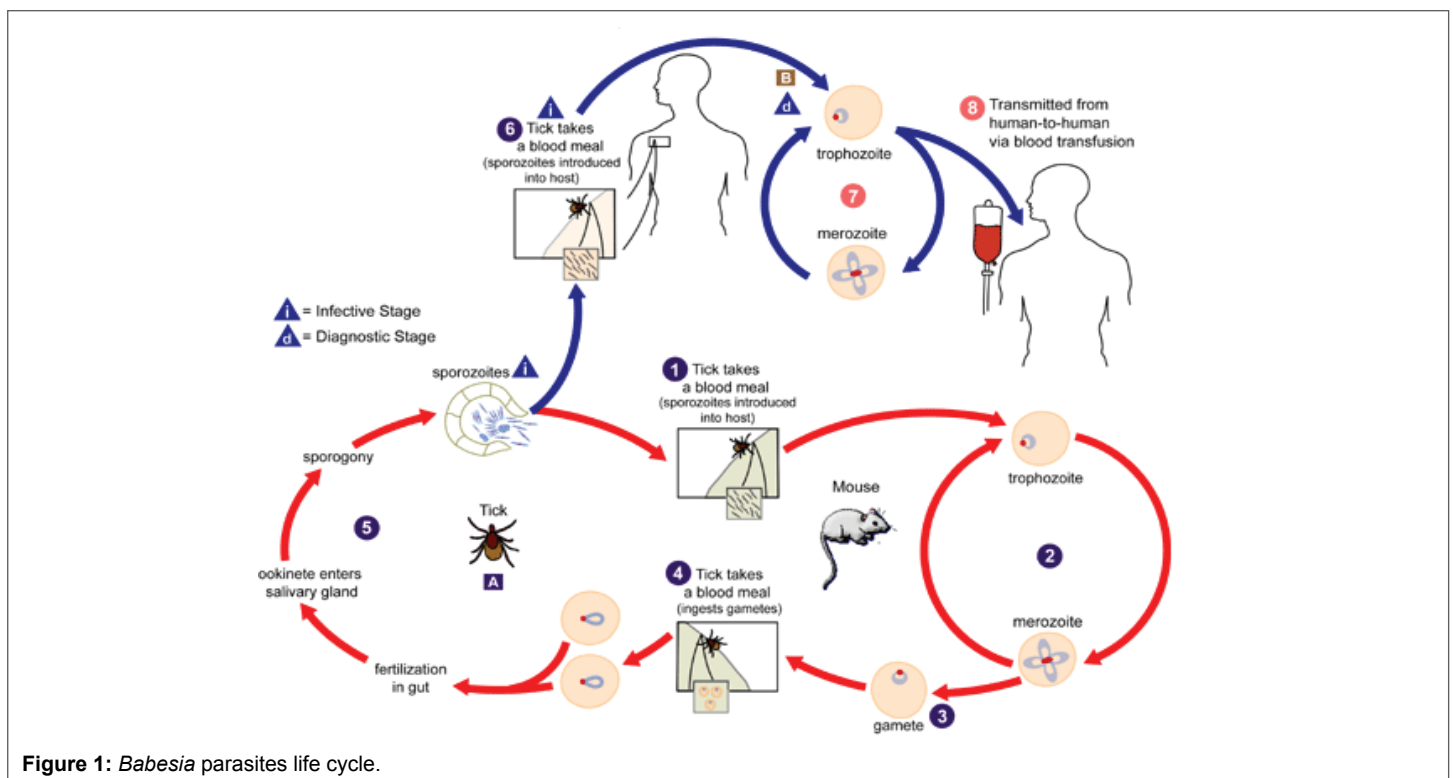


Figure 1: *Babesia* parasites life cycle.

*Babesia divergens* is the babesiosis cause, especially in Europe, and the transmission takes place *via Ixodes ricinus*, a tick with a three-year life cycle (larva, nymph, and adult). Most infections coincide with warmer weather (period of greater tick activity) and because of the higher number of people in areas of infestation [20].

Because of their peripheral position on the red blood cells, the parasites may be frequently confused with *Plasmodium falciparum* [13]. However, the species that cause babesiosis lack a tissue stage and rarely trigger large scale hemolysis, since their asexual reproduction is asynchronous [13,21].

## Epidemiology

Babesiosis is an emerging disease, widely distributed throughout the world, affecting several species of vertebrates, with the greatest impact on bovines and humans. Although humans are considered accidental hosts, the infections rates of this disease have increased considerable over the years [13,24,25]. In this sense, knowledge of the etiological agent, the vector, and the clinical characteristics are important for (1) clinical recognition of the disease (especially in terms of traveler's health), (2) correct diagnostic approach, and (3) appropriate therapeutic management.

The disease is considered endemic in the US and in Europe; however, there are reports of occurrence in other regions. One of the more recent studies demonstrated 48 cases in China, most among women with an average age of 45 year, and the infection was caused by *B. venatorum* [4]. Obviously, these reports do not compare to those registered in the US, where from 2005-2010 more than 1400 cases of human babesiosis were reported [8], and in 2013, 1762 cases were reported in 27 states [9]. Because of its geographical distribution and the annual number of cases, babesiosis caused by the agent *B. microti* is currently considered to be an emerging disease, and the cases involving *B. divergens* are rare [13]. In the US, the zoonosis involves rodents of the *Peromyscus leucopus* species as reservoirs and intermediate hosts for the pathogen [13]. In Europe, the diseases more rare, but also more lethal, the most caused by *B. divergens* in splenectomized patients [9].

The description of babesiosis in humans on the European continent includes countries such as Scotland and Russia – especially associated with cattle rearing [2,13,26,27], in addition to Germany, Austria, Spain, Ireland, Italy, Switzerland, Sweden, and especially France and England [17]. Although sporadic, the disease also occurs in countries in Asia including South Korea, Japan, and Taiwan, and also in countries of the African continent [13]. The first case of babesiosis in Australia was reported in 2012, after microscopic examination of the blood sample from a 56 year old, who died in 2011. The samples demonstrated the presence of intra-erythrocyte parasites [3]. Other human infections caused by *Babesia* spp, also occurred in South Africa, in Egypt, and in Mexico, which may mean that this parasite has a much wider distribution than is currently known [17].

In Brazil, one of the cases of human babesiosis, whose diagnosis was not conclusive during the course of the illness with relapses, was identified in the city of Rio de Janeiro (RJ), and the patient subsequently died. This was a case of a splenectomized patient [8]. Brazil has also been cited as origin of a case of babesiosis, described in Poland in 1997 [28].

## Immunology & Pathogenesis

Babesiosis is often compared to malaria in terms of physiopathogenesis, due to the intra-erythrocyte location of the agents [13,29,30]. In fact, erythrocyte invasion is an important part of the *Babesia* spp. cycle, and it is very efficient at entering these cells in order to evade the human host response. Interaction between this pathogen and the host cell is very specific, since invasion occurs only in red blood cells, a fact attributed to the merozoites that are capable of locating, binding, and invading them [31].

This specificity is possible due to the multiple adhesive interactions that occur between the ligands present on the protozoan membranes and the target receptors the surface of the host cell [32]. In *B. bovis*, for instance, several genes code for merozoite surface antigens (MSAs), specifically MSA-1, MSA-2b, and MSA-2c [33]. This genetic diversity may be strategic to the survival of these protozoa [34], and the antigenic variations arise from these diversity profiles [33].

The host immune response is required for control of the infection caused by *Babesia* spp., important in this process are the cytokines, highlighting that the sequential expression of their genes can confer protective immunity [35]. Studies have also suggested action of the TCD<sup>+</sup> lymphocytes and natural killer cells in response to the disease [36]. In response to infection, there is leukocyte activation, production of tumor necrosis factor-alpha (TNF- $\alpha$ ), and of interleukin 1 (IL-1) interleukin 2 (IL-2), and interleukin 6 (IL-6) [13,36], together with increasing levels of TNF which are, however, lower than those reached in malaria [37]. There is also lymphocyte activation and hypergamma globulinemia [38]. Specifically for *B. microti*, during the acute phase of infection, levels of IL-12, IFN- $\gamma$ , and TNF- $\alpha$  are elevated and are crucial to control the initial explosion of parasite multiplication. In these patients, the most common complication is lung infection, with detection of IFN- $\gamma$  within and around pulmonary blood vessels and TNF- $\alpha$  in the alveolar septum [35].

The function of the spleen in the disease pathogenesis is not completely established; however, the most severe form has been observed among patients with functional or anatomic asplenia. In part, the splenomegaly is associated with proliferation of phagocytic cells [13,39]. It is thought that the red blood cells infected by *B. divergens* alter spleen histopathology and cause alterations in the cell cycle and induce oxidative stress in this tissue [40].

Merozoites exit the red blood cells by rupture of the host cell, resulting in hemoglobin release in the blood stream. The free hemoglobin is immediately bound to haptoglobin and processed by phagocytic cells [13].

## Clinical Aspects

In general, the incubation period for babesiosis varies from one to three weeks, though it can extend to six weeks after the tick bite, or even up to nine weeks in case of infection by blood transfusion. On average, the time between symptom onset and diagnosis is 15 days. Among the cases verified in the United States and in Europe, the clinical manifestations differ markedly, considering that the infections are caused by different parasites [13].

The severity of the infection may vary with age and host immune status, simultaneously with other pathogens, and/or genetic factors. Acute infections in asplenic or immunocompromised patients, newborns, and the elderly, may give rise to complications, among which are: severe hemolysis, hemodynamic instability, acute respiratory failure, and multiple organ dysfunction syndrome, which may be fatal [41].

Symptoms are initially non-specific: malaise, fatigue, anorexia, fever (temperature above 38.0 °C and relative bradycardia), headache, chills, joint and muscle pain, and behavioral changes with emotional lability or depression [13,41]. Fever, important on physical examination, may be constant or intermittent. Other symptoms cover the progression of the disease with acute respiratory distress syndrome (ARDS) [42], shock, petechia, and other hemorrhagic phenomena, and hepatosplenomegaly. Association with Lyme disease is not uncommon [13]. Also common are: discrete hepatomegaly and splenomegaly, and absence of lymphadenopathy. On computerized tomography, splenic infarcts and ruptures have been observed in the absence of palpable splenomegaly [13]. However, asymptomatic infection has been registered in roughly 20% of adult patients and 50% of pediatric patients, in a study spanning one decade in a highly endemic area [14].

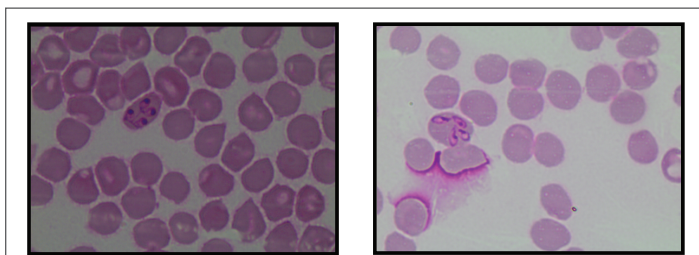
In Europe – where most cases are caused by *B. divergens* – evolution is fulminant, with fever, prostration, anemia, and hemoglobinuria, after the three week incubation period. Hemoglobinuria is present, followed by jaundice, persistent fever (between 40.0 to 41.0°C), headache, chills, sudoresis, and myalgia [13,41]. In contrast, in the US, most infections registered are asymptomatic or mild, and in 30% of cases is related to asplenia, with a low mortality rate. The incidence of subclinical infections is high, however, the clinical alterations are more common in asplenic patients, patients with Lyme disease (*Ixodes scapularis*, also the vector that transmits Lyme borreliosis), the elderly, and the predisposed, such as HIV-positive patients and those with lymphoma [13,43-45]. *Borrelia burgdorferi* and *Babesia microti* infections are common and can cause more severe human disease [46]. The convalescence period usually lasts a few weeks, but can extend as long as eighteen months [38,47].

## Laboratory Diagnosis

Diagnosis of babesiosis is usually established by blood smear, with Giemsa (or a derivative) staining [48], or by thick blood smear [13]. Detection of the pathogen, however, is obtained by visualization of the characteristic intra erythrocytic ring [47] (Figure 2), and the morphological similarity of these rings with those of *Plasmodium* spp. must be taken into account [20]. This fact highlights the importance of patient history, especially travel history to areas considered endemic [14]. However, confirmative tests based on molecular or serologic methods may confirm diagnosis [48], including in situations of low parasitemia [9]. Alternatively, in these situations, multiple smear tests should be performed over several days to confirm diagnosis [13].

The parasitemia of *Babesia* may also be obtained by inoculation of suspected patient's blood into laboratory animals, easily detected when the parasite is *B. microti* [49]. Although the majority of routine laboratories do not have adequate structures for use of molecular and serological methods for rapid and reliable detection of *Babesia*, they should be considered for the diagnosis of the disease [50]. Among the most widely known molecular approaches is polymerase chain reaction (PCR), used in situations where parasite detection is difficult, in patients presenting symptoms suggestive of the disease [13]. Upon using primers for amplification of specific gene regions such as 18S rRNA and the ITS (internal transcribed spacer) region and DNA extraction [51], the technique allows for differentiation of the *Babesia* spp. [13] with high specificity and sensitivity [49]. Additionally, real-time PCR increases the detection limit considerably [14,52]. In addition to PCR, DNA sequency and DNA microarray systems have shown promising preliminary results with infected bovines [53].

The immunological diagnosis of babesiosis is normally established by indirect immunofluorescence (IFA). This is considered the standard assay for detection of *Babesia* spp. antibodies; however, other techniques such as Immunoblot, immunochromatography, and ELISA (enzyme-linked immunosorbent assay) are also used [49]. Specifically, in IFA when using *B. microti* as antigen, antibody detection reaches 88-96% in infected patients. The IFA slides containing the antigen are prepared using parasited erythrocytes produced in hamsters [9].



**Figure 2:** *Babesia* parasites in red blood cells on a stained blood smear (Aécio Carlos de Oliveira / UFV Veterinary Department).

Laboratory alterations include hemolytic anemia with increased reticulocytes, and decreased haptoglobin [13], followed by increased blood sedimentation rate, thrombocytopenia, leucopenia (with lymphopenia), proteinuria, hemoglobinuria, and elevated liver enzymes (aminotransferases, alkaline phosphatase, and lactate dehydrogenase) [13,54]. Specifically, the infections caused by *B. microti* are accompanied by decreased hematocrit and hemoglobin counts, elevated total bilirrubins, reduced haptoglobin, and increased reticulocytes, more consistent with hemolytic anemia. Infections by *B. divergens* show mainly reduced hemoglobin levels (4 to 8 g/dL), indicative of intense hemolysis [13].

## Treatment

Treatment of babesiosis – in symptomatic patients – is carried out with antimicrobial therapy, after diagnostic confirmation [35]. The most commonly used antibiotics are: (1) atovaquone, azithromycin, and a combination of the two; and (2) clindamycin, quinine, and a combination of the two. The binomial dose/duration of administration of these agents generally varies between adult and pediatric patients [14].

In moderate cases of babesiosis caused by *B. divergens* and *B. venatorum*, the use of clindamycin for seven to ten days is widely adopted; in infections caused by *B. microti* the combination of atovaquone and azithromycin is indicated, for the same treatment period [20]. These antibiotics and their combinations are generally administered orally. The most cited side effects are diarrhea and exanthema (azithromycin and atovaquone). It should be highlighted that due to problems attributed to resistance to the principal antimicrobial regimens, the combinations of quinine and clindamycin or atovaquone and azithromycin have been used [12]. However, while treatment with these combinations is effective, there are reports of problems with parasite persistence and response time in endemic areas. Although resistance to these drugs is not suspected in these cases, they do highlight the need for ongoing monitoring of parasitemia during patient treatment [20].

Asymptomatic patients infected with *B. microti* do not require treatment, unless species of *Babesia* spp. are detected by blood smear or PCR over more than three months. Symptomatic patients should not be treated if the blood smear or PCR are negative. Treatment is only recommended in case of pathogen detection [13, 25].

In the United States, patients without risk factors or baseline disease usually present low parasitemia, and recovery in most cases without need of specific chemotherapy [37]. More severely affected patients are generally treated with clindamycin 300-600 mg four times daily (by intravenous route), associated with quinine 650 mg thrice daily (by oral route) for seven to ten days. Moreover, some critical patients may require supportive treatment, including: antipyretics, vasopressors, blood transfusion and mechanical ventilation [9].

Chemotherapy and blood transfusions are recommended in Europe and in the more severe cases in the US. The objective of these procedures is to ameliorate parasitemia levels [13,55].

## Prophylaxis and Control

The best preventive measure for babesiosis is avoiding exposure to the habitats of the tick disease vectors, especially in endemic areas and between the months of May and September [13]. Specifically in the case of *Ixodes scapularis*, they are most commonly found in areas with shrubs and grasses according to the cycle of this vector [9], and activities in such locations require attention.

To date, no vaccine is available, however, efforts have been made to produce one. Despite this, simple efforts help in preventing tick bites and the infections they transmit [9].

In situations of absolute need to visit exposure risk areas, long-sleeves, pants, and socks should be worn, and insect repellents used, especially those containing permethrin (on the clothes and DEET (diethyltoluamide) on the skin [14]. Wearing light colored clothing facilitates visualization and subsequent removal of the ticks [9]. In these areas, care should be taken to walk on open trails, remaining in the center of the path and avoiding exposure to grasses where there is a higher concentration of ticks [9].

Reduction of transmission *via* blood transfusion can be achieved by exclusion of donors from endemic areas or those with a recent history of tick bites (within two months of donation) [56]. The possibility of babesiosis should always be remembered upon differential diagnosis of transfusion transmitted infections.

### Final Considerations

The emergence of babesiosis together with the cosmopolitan distribution of ticks and the environmental changes that favor their cycle are factors that have elevated the zoonotic potential of the disease. New species of *Babesia* spp. have been described in distinct geographical regions and demonstrate the potential for transmission to humans [57]. The increased diversity and incidence of infections over the recent years has been attributed to better diagnosis, greater awareness on the part of the public, greater contact with natural areas and consequently with the vectors, as well as to environmental changes and the increased number of immunosuppressed individuals [15]. Autoimmune hemolytic anemia (AIHA) may be triggered by babesiosis [58].

Distributed worldwide, little is known about the prevalence of *Babesia* in countries where malaria is endemic, because of the difficulty in distinguishing it from *Plasmodium*. There are differences in the species that cause babesiosis, in relation to the regions where the disease occurs.

*B. divergens* is the cause of most cases on the European continent and in the US, *B. microti* is the principal cause, especially in the northeast and Midwest regions [9].

In Brazil the cases are more restricted, however, another form of transmission – which occurs among travelers – merits attention, given the increased number of travels over the last decades. In this context, information about the transmissible illnesses at the destination is important. Specifically, prevention of babesiosis requires care in endemic areas. In terms of patient care, it is also important to carefully identify the causes of the disease presented, including knowledge of the natural history and place of travel, in order to precisely carry out anamnesis and physical exam [59,60]. Thus it is imperative to know the natural history of the vectors and the species of *Babesia* that infect humans [15].

Thus, care with education and training of health professionals in relation to this zoonosis is becoming essential, and is fundamentally important in recognizing the disease. Outbreaks may occur as a result of the pathogen's adaptability to its hosts and climate, which also highlights the importance of the measures mentioned above. Familiarity with the symptoms triggered by babesiosis together with knowledge of efficient diagnostic methods for confirmation is fundamental to establish early treatment and the necessary prophylactic measures.

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### References

1. Ríos L, Alvarez G, Blair S (2003) Serological and parasitological study and report of the first case of human babesiosis in Colombia. *Revista da Sociedade Brasileira de Medicina Tropical* 36: 493-498.
2. Stafford III KC, Williams SC, Magnarelli LA, Bharadwaj A, Ertel SH, et al. (2014) Expansion of zoonotic babesiosis and reported human cases, Connecticut, 2001-2010. *J Med Entomol* 51: 245-252.
3. Papparini A, Senanayake SN, Ryan UM, Irwin PJ (2014) Molecular confirmation of the first autochthonous case of human babesiosis in Australia using a novel primer set for the beta-tubulin gene. *Exp Parasitol* 141: 93-97.
4. Jiang JF, Zheng YC, Jiang RR, Li H, Huo QB, et al. (2015) Epidemiological, clinical, and laboratory characteristics of 48 cases of "*Babesia venatorum*" infection in China: a descriptive study. *Lancet Infect Dis* 15: 196-203.
5. Skrabalo Z, Deanovic Z (1957) Piroplasmosis in man; report of a case. *Doc Med Geogr Trop* 9: 11-16.
6. Alecrim I, Pinto B, Ávila T, Costa R, Pessoa I (1983) Registro do primeiro caso de infecção humana por *Babesia* spp no Brasil. *Rev Patol Trop* 12: 11-29.
7. Gazeta GS, Carvalho RW, Avelar RF, Amorim M, Aboud-Dutra AE (2004) Brazilian Archive of Veterinary Medicine and Zootechnics 56: 741-744.
8. Serra-Freire NM (2014) Case Index of Human Babesiosis in Rio De Janeiro. *Brazil Revista Uniabeu* 7: 15.
9. CDC (2015) Parasites -*Babesia microti*. Centers for Disease Control and Prevention, US.
10. Tome ACN, Canello TB, Luna EJA, Andrade Jr HF (2013) Health problems awareness during travel among faculty members of a large university in Latin America: preliminary report. *Rev Inst Med Trop Sao Paulo* 55: 55-59.
11. Matos V, Barcellos C (2010) Relações entre turismo e saúde: abordagens metodológicas e propostas de ação. *Revista Panamericana de Salud Pública* 28: 128-134.
12. Hildebrandt A, Gray JS, Hunfeld KP (2013) Human babesiosis in Europe: what clinicians need to know. *Infection* 41: 1057-1072.
13. Gelfand JA, Vannier EG (2010) *Babesia* Species. In: Mandell GL, Bennett JE, Dolin R (eds) Principles and practice of infectious diseases. 7<sup>th</sup> edition, Elsevier, Philadelphia, US.
14. Vannier EG, Diuk-Wasser MA, Mamoun CB, Krause PJ (2015) Babesiosis. *Infect Dis Clin North Am* 29: 357-370.
15. Yabsley MJ, Shock BC (2013) Natural history of Zoonotic Babesia: role of wildlife reservoirs. *Int J Parasitol Parasites Wildl* 2: 18-31.
16. Herwaldt BL, Cacció S, Gherlinzoni F, Aspöck H, Slemenda SB (2003) Molecular characterization of a non-*Babesia divergens* organism causing zoonotic babesiosis in Europe. *Emerg Infect Dis* 9: 943-948.
17. Leiby DA (2011) Transfusion-transmitted *Babesia* spp.: bull's-eye on *Babesia microti*. *Clin Microbiol Rev* 24: 14-28.
18. Foppa IM, Krause PJ, Goethert H, Gem L, Brand B, et al. (2002) Entomologic and serologic evidence of zoonotic transmission of *Babesia microti*, eastern Switzerland 8: 722-726.
19. Lobo CA, Rodriguez M, Cursino-Santos JR (2012) *Babesia* and red cell invasion. *Current opinion in hematology* 19: 170-175.
20. Ord RL, Lobo CA (2015) Human Babesiosis: Pathogens, Prevalence, Diagnosis, and Treatment. *Current clinical microbiology reports* 2: 173-181.
21. Chiodini PL (1996) Babesiosis. In: Manson-Bahr PEC, Apter FIC (eds) Manson's Tropical Diseases. 10<sup>th</sup> edition, Ballière Tindall, London, UK.
22. Beaver PC, Jung RC, Cupp EW (1984) *Clinical Parasitology*. Lea and Febiger, Philadelphia.
23. Schleupner CJ (1996) Nosocomial infections associated with transfusion of blood and blood products. In: Mayhall CG (eds) Hospital Epidemiology and Infection Control. 1<sup>st</sup> edition, Williams & Wilkins, Baltimore, US.

24. Mosqueda J, Olieria-Ramírez A, Aguilar-Tipacamú G, Cantó GJ (2012) Current advances in detection and treatment of babesiosis. *Curr Med Chem* 19: 1504-1518.
25. Gonzalez LM, Rojo S, Gonzalez-Camacho F, Luque D, Lobo CA, et al. (2014) Severe babesiosis immunocompetent man, Spain, 2011. *Emerging infectious diseases* 20: 724-726.
26. Ruebush TK, Juranek DD, Spielman A, Piesman J, Healy GR (1981) Epidemiology of human babesiosis on Nantucket Island. *Am J Trop Med Hyg* 30: 937-941.
27. Spielman A, Wilson ML, Levine JF, Piesman J (1985) Ecology of ixodes dammini-borne human babesiosis and Lyme disease. *Annu Rev Entomol* 30: 439-460.
28. Humiczewska M, Kuźna-Grygiel W (1997) A case of imported human babesiosis in Poland. *Wiad Parazytol* 43: 227-229.
29. Allred DR (1998) Antigenic variation in *Babesia bovis*: how similar is it to that in *Plasmodium falciparum*? *Ann Trop Med Parasitol* 92: 461-472.
30. Clark IA, Jacobson LS (1998) Do babesiosis and malaria share a common disease process? *Ann Trop Med Parasitol* 92: 483-488.
31. Lobo CA (2005) *Babesia divergens* and *Plasmodium falciparum* use common receptors, glycoporphins A and B, to invade the human red blood cell. *Infect Immun* 73: 649-651.
32. Yokoyama N, Okamura M, Igarashi I (2006) Erythrocyte invasion by *Babesia* parasites: current advances in the elucidation of the molecular interactions between the protozoan ligands and host receptors in the invasion stage. *Vet Parasitol* 138: 22-32.
33. Yokoyama N, Sivakumar T, Tuvshintulga B, Hayashida K, Igarashi I, et al. (2015) Genetic variations in merozoite surface antigen genes of *Babesia bovis* detected in Vietnamese cattle and water buffaloes. *Infect Genet Evol* 30: 288-295.
34. Deitsch KW, Lukehart SA, Stringer JR (2009) Common strategies for antigenic variation by bacterial, fungal and protozoan pathogens. *Nat Rev Microbiol* 7: 493-503.
35. Vannier E, Gewurz BE, Krause PJ (2008) Human babesiosis. *Infect Dis Clin North Am* 22: 469-488.
36. Shaio MF, Lin PR (1998) A case study of cytokine profiles in acute human babesiosis. *Am J Trop Med Hyg* 58: 335-337.
37. Gelfand JA (1995) *Babesia*. In: Mandell GL, Bennett JE & Dolin R (eds) *Principles and Practice of Infectious Diseases*. 4th edition, Elsevier Health Sciences, Amsterdam, Netherlands.
38. Benach JL, Habicht GS, Hamburger MI (1982) Immuno responsiveness in acute babesiosis in humans. *J Infect Dis* 146: 369-380.
39. Wiercinska-Drapalo A, Grzeszczuk A, Panasiuk A, Prokopowicz D (1998) Babesiosis-disease of human and animals. *Pol Arch Med Wewn* 99: 239-244.
40. Dkhil MA, Al-Quraishy S, Al-Khalifa MS (2014) The Effect of *Babesia divergens* Infection on the Spleen of Mongolian Gerbils. *BioMed research international* 2014: 1-8.
41. Schnittger L, Rodriguez AE, Florin-Christensen M, Morrison DA (2012) *Babesia*: A world emerging. *Infection, Genetics and evolution* 12: 1788-1809.
42. Hemmer RM, Wozniak EJ, Lowenstine LJ, Plopper CG, Wong V, et al. (1999) Endothelial cell changes are associated with pulmonary edema and respiratory distress in mice infected with the WA1 human *Babesia* parasite. *J Parasitol* 85: 479-489.
43. Benezra D, Brown AE, Polsky B, Gold JW, Armstrong D (1987) Babesiosis and infection with human immunodeficiency virus (HIV). *Ann Intern Med* 107: 944.
44. Gorenflot A, Moubri K, Precigout E, Carcy B, Schetters TP (1998) Human babesiosis. *Annals of Tropical Medicine and Parasitology* 92: 489-501.
45. Machtinger L, Telford SR, Inducil C, Klapper E, Pepkowitz SH, et al. (1993) Treatment of babesiosis by red blood cell exchange in an HIV-positive splenectomized patient. *Journal of Clinical Apheresis* 8: 78-81.
46. Diuk-Wasser MA, Edouard V, Peter JK (2016) Coinfection by *Ixodes* tick-borne pathogens: Ecological, epidemiological, and clinical consequences. *Trends parasitol* 32: 30-42.
47. Gelfand JA, Callahan MV (1998) Babesiosis. *Current Clinical Topics in Infectious Diseases* 18: 201-216.
48. Mørch K, Holmaas G, Frolander PS, Kristoffersen EK (2015) Severe human *Babesia divergens* infection in Norway. *International Journal of Infectious Diseases* 33: 37-38.
49. Vannier E, Krause PJ (2009) Update on babesiosis. *Interdisciplinary perspectives on infectious diseases* 2009: 984568.
50. Zhou X, Xia S, Huang JL, Tambo E, Ge HX, et al. (2014) Human babesiosis, an emerging tick-borne disease in the People's Republic of China. *Parasites and Vectors* 7: 1-10.
51. Blaschitz M, Narodoslavsky-Gföller M, Kanzler M, Stanek G, Walochnik J (2008) *Babesia* species occurring in Austrian *Ixodes ricinus* ticks. *Applied and environmental microbiology* 74: 4841-4846.
52. Akel T, Neville M (2017) Hematologic manifestations of babesiosis. *Annals of clinical and Laboratory Research* 5: 1-7.
53. El-Ashker M, Hotzel H, Gwida M, El-Beskawy M, Silaghi C, et al. (2014) Molecular biological identification of *Babesia*, *Theileria*, and *Anaplasma* species in cattle in Egypt using PCR assays, gene sequence analysis and a novel DNA microarray. *Vet parasitol* 207: 329-334.
54. Kim N, Rosenbaum GS, Cunha BA (1998) Relative bradycardia and lymphopenia in patients with babesiosis. *Clin Infect Dis* 26: 1218-1219.
55. Evenson DA, Perry E, Kloster B, Hurley R, Stroncek DF (1998) Therapeutic apheresis for babesiosis. *J Clin Apher* 13: 32-36.
56. Krause PJ, Spielman A, Telford SR, Sikand VK, Mc Kay K, et al. (1998) Persistent parasitemia after acute babesiosis. *N Engl J Med* 339: 160-165.
57. Hunfeld KP, Hildebrandt A, Gray JS (2008) Babesiosis: recent insights into an ancient disease. *International journal for parasitology* 38: 1219-1237.
58. Narurkar R, Mamorska-Dyga A, Nelson JC, Liu D (2017) Autoimmune hemolytic anemia associated with babesiosis. *Biomarker Research* 5: 1-4.
59. Daly R, Chiodini P (2008) Laboratory Investigations and Diagnosis of Tropical Diseases in travelers. *Infect Dis Clin North Am* 26: 803-818.
60. Cruz AT (2013) Infections in returned travelers. *Clinical Pediatric Emergency Medicine* 14: 118-134.